Note

Sophorose produced in the fusion and Koenigs-Knorr reactions

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Sophorose (1, 2-O- β -D-glucopyranosyl-D-glucose) has been produced via the fusion reaction of 1,3,4,6-tetra-O-acetyl- α -D-glucopyranose (2) in the presence of toluene-p-sulphonic acid¹, and by the Koenigs-Knorr reaction of 2,3,4,6-tetra-O-acetyl- α -D-glucopyranosyl bromide (3) and methyl 4,6-O-benzylidene- α -D-glucopyranoside^{2,3}. In contrast, kojibiose (4, 2-O- α -D-glucopyranosyl-D-glucose) is produced in the fusion reaction of 3,4,6-tri-O-acetyl-1,2-anhydro- α -D-glucopyranose (5) in the absence of catalyst⁴, and in the Koenigs-Knorr reaction of 2 and 3^{5,6}. A mixture of 1 and 4 was produced in the fusion reaction of 2 with ZnCl₂ as catalyst^{1,7}.

We now report the formation of sophorose (1) via the fusion reaction of 5, or of a mixture of 2 and 5, in the presence of toluene-p-sulphonic acid, and in the Koenigs-Knorr reaction of 2 and 3.

Compound 2, 5, or a mixture of 2 and 5 melted at $65-130^{\circ}$ in the presence of toluene-p-sulphonic acid or $ZnCl_2$ (Table I). The reaction products were deacetylated, and separated into three fractions by chromatography on Bio-Gel P-2. A series of oligo- and poly-saccharides was detected by p.c. (R_{Gle} 0.54, 0.33, 0.26, 0.20, and slowly moving spots; solvent A). Essentially the same products were formed in these reactions, but the fusion of 5 in the presence of $ZnCl_2$ produced only a trace of oligo- and poly-saccharides (p.c.), and 2 did not produce 5. The oligo- and poly-saccharide fraction was obtained in up to 38.6% yield; reaction D gave a slightly higher yield than reactions A-C (Table I).

The product having R_{Gle} 0.54, isolated by preparative p.c. as a syrup from reactions A-D in yields of 19, 16, 13, and 20%, respectively, was a mixture of 1 and 4. It had a reducing value of 45-55% compared with D-glucose, and exhaustive digestion with β -D-glucosidase gave a mixture of glucose and disaccharide 4 (R_{Gle} 0.54; reducing value, 52%). The product from reaction $C^{1,7}$ contained a significant amount of 4, but only traces were present in the corresponding products from reactions A, B^1 , and D.

The product $(R_{Glc} \ 0.54)$ from reaction D gave a crystalline octa-acetate monohydrate with R_F values [t.l.c.; 0.63 (solvent C), 0.59 (solvent D)] indistinguishable from those of the octa-acetates of 1 and 4. The octa-acetate monohydrate had $[\alpha]_D^{20}$

TABLE I

OLIGO- AND POLY-SACCHARIDES PRODUCED IN THE FUSION REACTION

Reactiv	Reaction Monosacchande	Catalyst ^b	Reaction		Oligo- and	Oligo- and poly-saccharides	S	,	
	derivative ^a		Temperature (degrees)	Time (mm)	Fraction	Yield (%)	D.p.c	Anthrone value ^{it} (%)	[a] ^{1,} (c I, water) (degrees)
A	w	TsOH	65 ± 3	10	с.	trace	n.d.	n.d.	n.d.
Ве	2	TsOH	105 ± 3	30	ಎ ಇ -	5 co 5	5.0 5.2 5.4	8 8 8	- 1 - - 1 - 1
ర	2	ZnCl2	105 ± 3	09	2 65 4	9 - 6	11	8 8 8	- 1 + 2 7.5 3 6 f
Q	2 + 5	ТѕОН	130 ± 3	40	2 a 2	7.3	10 3.0	8 33	+
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"2, 1,3,4,6-Tetra-O-acetyl-α-D-glucopyranose; 5, 3,4,6-t11-O-acetyl-1,2-anhydro-α-D-glucopyranose. ¹TsOH, p-Toluenesulfonic acid. ²Degree of polymerization. ²Calc. as D-glucose. ²Similar teactions were reported in Refs. 1 and 7, ⁷α-D-Glucoside was present (Refs. 1 and 7).

NOTE 149

+42° (c 0.55, chloroform) (cf. +45° for the anhydrous compound¹³), showed eight signals for acetate groups in the n.m.r. spectrum (CDCl₃), and had $v_{\text{max}}^{\text{KBr}}$ at 890 cm⁻¹ characteristic of a β -D-glucopyranoside. The deacetylated product gave only D-glucose (p.c.) on complete hydrolysis (2m HCl, 100°, 3 h), had a reducing value of 48% compared with D-glucose, and was completely hydrolysed by β -D-glucosidase.

The other oligo- and poly-saccharides were not further fractionated. Fraction a consumed 0.9-1.1 mol of periodate per D-glucose residue, and formic acid could not be detected by n.m.r. spectroscopy¹². The $[\alpha]_D$ value $(-3^{\circ}$ to -7.5°) accorded with that (-8°) of $(1\rightarrow 2)$ - β -D-glucan⁸.

The products which were obtained by the Koenigs-Knorr reaction of 2 and 3 in nitromethane^{5,6} in the presence of $Hg(CN)_2$, after deacetylation, gave 52% of a syrupy disaccharide fraction which had chromatographic behaviour identical with that of the disaccharide fraction obtained from reaction C above. Exhaustive digestion with β -D-glucosidase gave D-glucose and 4 (detected by t.l.c.), and indicated that 1 and 4 (\sim 3:2) were present in the disaccharide fraction. Compound 4 was previously isolated in 21% yield from the same reaction^{5,6}.

EXPERIMENTAL

Compounds 2^5 , 3^9 , and 5^{10} were prepared by the conventional methods. Almond β -D-glucosidase (EC 3.2.1.21) was obtained from Sigma (Lot. 29B-1280); the enzyme (1 mg) liberated ~2.5 μ mol of D-glucose per min from salicin at pH 5.2 and 37°.

P.c. was performed by the descending method on Whatman No. 1 or 3MM filter papers with A, acetic acid-1-butanol-ethanol-water (12:50:5:25), B, 1-butanol-pyridine-water (3:2:1:5), and detection with alkaline silver nitrate. T.l.c. was performed on Silica Gel G (Merck, Type 60) with C, benzene-ethyl acetate (4:3), D, benzene-acetone (4:1), and detection by charring with sulphuric acid. The reducing-sugar values were obtained by the Schales method¹¹, and the degree of polymerisation (d.p.) was calculated therefrom. The other methods were described previously¹².

Fusion reactions. — The reactions A-D were performed by using 1-3 g of 2 or 5 (Table I). The products were deacetylated with methanolic ammonia and eluted from a column (1.7 × 213 cm) of Bio-Gel P-2 with 10% ethanol (7-ml fractions). Fractions were monitored by the anthrone reaction and by p.c., and grouped as follows: a (fractions 20-30) contained products having low mobility, and $R_{\rm Glc}$ 0.20 and 0.26 (solvent A); b (fractions 31-43), $R_{\rm Glc}$ 0.26, 0.33, 0.54, and 1.00 (trace); and c (fractions 44-58), $R_{\rm Glc}$ 0.54 (trace) and 1.00. Fractions b and c were rechromatographed to remove monosaccharide, concentrated in vacuo at 40° to a small volume, and lyophilized. All the products were amorphous and soluble in water.

Fraction **b** (277 mg) from reaction D was subjected to preparative p.c. on three sheets (46 \times 57 cm) of Whatman 3MM paper, and the compound having $R_{\rm Glc}$ 0.54 was eluted from the appropriate areas with hot water. The extract was concen-

150 NOTE

trated *in vacuo*, and the resulting syrup (184 mg), $[\alpha]_D^{15} + 20^\circ$ (c 1, water), was treated with acetic anhydride and pyridine. Crystallisation of the product from ether-light petroleum (b.p. 40-60°) gave the octa-acetate (78%), m.p. 74-76°, $[\alpha]_D^{21} + 42^\circ$ (c 0.55, chloroform); $v_{\text{max}}^{\text{KBr}}$ 1760 (acetyl C=O) and 1240 (acetyl C-O), 1090-1040 (C-O), 890 cm⁻¹ (β -D-Glc).

Anal. Calc. for C₂₈H₃₈O₁₉ · H₂O: C, 48.27; H, 5.79. Found: C, 48.61; H, 5.60. The octa-acetate (1 mg) was treated with 0.2m NaOH in ethanol (0.2 ml) at room temperature overnight. Ethanol was evaporated in vacuo, and the residue was made up to 5 ml with 0.2m acetate buffer (pH 5.2). The reducing-sugar value was 44% against D-glucose. β-D-Glucosidase (0.1 mg) was added to the solution, and the mixture was incubated at 37°. Aliquots (0.5 ml) were withdrawn at intervals, and the reducing-sugar value was determined. The value reached 98% against D-glucose after incubation for 2 h.

The Koenigs-Knorr reaction. — To a solution of 2 (1.88 g) in nitromethane (40 ml) was added $Hg(CN)_2$ (2 g) with stirring. To the mixture was added a solution of 3 (2.23 g) in anhydrous chloroform (10 ml), and stirring was continued for 24 h in the dark. More 3 (1.6 g × 3) was added, and 2 disappeared (t.l.c.) after 72 h. The mixture was filtered, and concentrated to dryness. The residue was deacetylated with methanolic sodium methoxide, and the products were eluted from a column (3 × 60 cm) of Silica Gel 60 (Merck) with 1-butanol-ethyl acetate-2-propanol water (2:1:1:1) to remove monosaccharide. The disaccharide fraction was isolated as syrup (0.97 g), R_{Gle} 0.57 (p.c., solvent B). The octa-acetate had R_F 0.62 (solvent C) and 0.59 (solvent D) in t.l.c.

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